PROPAGATION OF *Orycles* NUDIVIRUS FOR THE MANAGEMENT OF RHINOCEROS BEETLE

RAMLE MOSLIM; NORMAN KAMARUDIN and SITI RAMLAH AHMAD ALI



MPOB INFORMATION SERIES • ISSN 1511-7871 • JUNE 2015

MPOB TS No. 146

hinoceros beetle, *Oryctes rhinoceros* (Coleoptera: Scarabaeidae) (*Figure 1*) is a major pest of the oil palm, causing 25% of crop losses two years after infestation (Norman *et al.*, 1997). The *Oryctes* nudivirus (OrNV) (*Figure 2*) is a biological control agent of the rhinoceros beetle (Huger, 1966). The OrNV was discovered in Malaysia and introduced into many coconut and oil palm producing countries to control the rhinoceros beetle (Bedford, 2014). In Malaysia, four types of OrNV



Figure 1. Rhinoceros beetle, Oryctes rhinoceros adult.

were identified where Type B was the most virulent compared to Types A, C and D, causing 65% mortality on larvae and 86.7% on adults (*Figure 3*) (Ramle *et al.*, 2005). Introduction of OrNV Type B to oil palm plantations has reduced the population of rhinoceros beetle and palm damage (Ramle *et al.*, 2011).

INSECT CELL CULTURE

The insect cell line DSIR-HA-1179 (*Figure 4*) was developed from the black beetle, *Heteronychus arator* (Coleoptera: Scarabaeidae) and used to propagate high purity OrNV (Crawford, 1982). The cells were propagated in a 25 cm² flask containing 5 ml of fresh prepared PS100 medium. The flasks were then placed inside an air tight container and stored in an incubator at 27° C for 15 days. The cell production reached up to 6.0×10^{5} cells/ml two weeks after incubation (*Figure 5*).

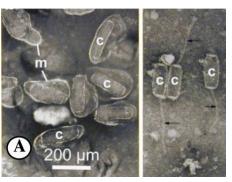


Photo by Huger et al. (2005)

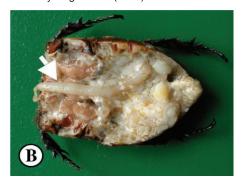


Figure 2. A) Morphological characteristics of Oryctes nudivirus, B) symptom of infected adult having swollen gut, full with milky content (arrow).

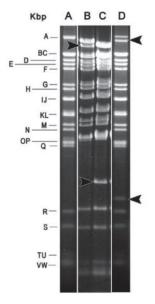
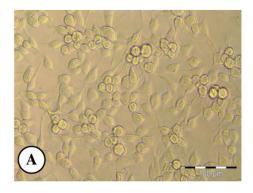


Figure 3. DNA profiles of four types of Oryctes nudivirus in Malaysia.







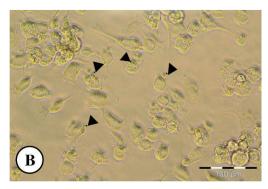


Figure 4. Morphological characteristics of insect cell line DSIR HA-1179. A) Healthy cells and B) nudivirus infected cells with cytophatic effect (arrows).







Figure 5. Some apparatus used for production of Oryctes nudivirus.

A) Incubator, B) 25 cm² cell culture flasks and C) microtubes containing formulated Oryctes nudivirus suspensions.

PRODUCTION OF NUDIVIRUS

- 1. Preparation of nudivirus suspension from nudivirus infected gut tissues using filtration techniques, via 0.45 μ m then 0.22 μ m membrane filter.
- 2. Adding virus suspension into a flask with insect cells between 60% to 70% confluent.
- 3. Incubate at 27°C for 15 days. Infected cells will show morphological changes or cytophatic effect.
- 4. Harvesting of nudivirus from the cell culture using centrifugation method.
- 5. Formulated nudivirus suspension in a 2.0 ml sterile microtube and store at 4°C.

 Conduct product quality analysis to identify type of nudivirus and determine the concentration of virus by Tissue Culture Infectious Dose 50% (TCID₅₀) analysis.

INTRODUCTION OF NUDIVIRUS IN THE FIELD

Three phases are involved (Ramle et al., 2003) as follows:

Phase 1: Pre-release Site Assessment

- 1. Identification of existing type of nudivirus.
- 2. Determination of nudivirus infection on immatures and adults *Oryctes*.

- 3. Assessment of palm damage.
- 4. Collection of young healthy adults for nudivirus inoculation and dissemination.

Phase 2: Time of Release

1. Inoculation of young adults and field release.

Phase 3: Post-release Monitoring and Impact Assessment

- 1. Determination of adult beetle population by trapping method.
- 2. Determination of virus infection on immatures and adults *Oryctes*.
- 3. Assessment of palm damage.
- 4. Determination on the spread of released virus.

BENEFITS

The cell culture could be used to propagate a specific OrNV which has high infectivity against identified beetle populations. Furthermore, it can be used to monitor the quality of individual OrNV prior to field release.

COST

Estimated cost to produce 50 ml of *Oryctes* nudivirus suspension in 2015 is RM 1000 (subject to change). This volume is sufficient for inoculating 1250 rhinoceros beetle adults. The cost includes training on introduction of nudivirus in the field using capture, inoculate and release technique.

REFERENCES

BEDFORD, G O (2014). Advances in the control of rhinoceros beetle, *Oryctes rhinoceros* in oil palm. *J. Oil Palm Res. Vol.* 26 No. 3: 183-194.

CRAWFORD, A M (1982). A coleopteran cell line derived from *Heteronychus arator* (Coleoptera: Scarabaeidae). In Vitro, 18: 813-816.

HUGER, A M (1966). A virus disease of the Indian rhinoceros beetle *Oryctes rhinoceros* (Linnaeus), caused by a new type of insect virus, *Rhabdionvirus oryctes* gen. n., sp. N. *J. Invertebrate Pathology*, 8: 38 – 51.

NORMAN, K and MOHD BASRI, W (1997). Status of rhinoceros beetle, *Oryctes rhinoceros* (Coleoptera: Scarabaeidae) as a pest of young oil palm in Malaysia. *The Planter*, 73(850):5-21.5.

RAMLE, M; NORMAN, K and MOHD BASRI, W (2003). *Oryctes* virus for biocontrol of rhinoceros beetle, *Oryctes rhinoceros*. *MPOB Information Series No.* 175: 4 pp.

RAMLE, M; WAHID, M W; NORMAN, K; GLARE, T G and JACKSON, T A (2005). The incidence and use of *Oryctes rhinoceros* virus for control of rhinoceros beetle in oil palm plantations in. *J. Invertebrate Pathology*, 89: 85-90.

RAMLE MOSLIM; NORMAN KAMARUDIN; ID-RIS A GHANI; MOHD BASRI WAHID; TREVOR A JACKSON; TEY, C C and MOHD AHDLY, A (2011). Molecular approaches in the assessment of *Oryctes rhinoceros* virus for the control of rhinoceros beetle in oil palm plantations. *J. Oil Palm Res. Vol.* 23: 1096-1109.

For more information, kindly contact:

Director-General MPOB 6 Persiaran Institusi, Bandar Baru Bangi, 43000 Kajang, Selangor, Malaysia. *Tel*: 03-8769 4400

Fax: 03-8925 9446 www.mpob.gov.my